

Separation of mycoplasma and acholeplasma from veggies.

Alexey Trushin*

Department of Biophysics, Federal University, Russia

Introduction

Vegetables are a crucial component of a healthy diet, providing essential nutrients and vitamins. However, they can sometimes harbor harmful microorganisms such as Mycoplasma and Acholeplasma, which pose potential health risks if consumed. These bacteria are often challenging to detect and remove due to their small size and elusive nature. Nevertheless, advancements in science and technology have enabled researchers to develop methods for effectively separating these pathogens from vegetables, ensuring food safety and consumer health [1, 2].

Mycoplasma and Acholeplasma are both members of the class Mollicutes, characterized by their lack of a cell wall. This unique feature makes them resistant to many conventional antibiotics and complicates their detection and eradication. While Mycoplasma species are associated with various infections in animals and humans, Acholeplasma species are primarily found in plants, including vegetables. The small size and morphological simplicity of Mycoplasma and Acholeplasma present significant challenges in separating them from vegetables [3, 4].

Conventional methods such as washing and surface sterilization may not effectively eliminate these bacteria, as they can adhere tightly to the vegetable surface or penetrate the plant tissue. Researchers have been exploring innovative techniques to overcome the challenges of separating Mycoplasma and Acholeplasma from vegetables. One promising approach involves the use of molecular biology tools such as polymerase chain reaction (PCR) and DNA sequencing to specifically identify and quantify these pathogens. By targeting unique genetic markers, scientists can distinguish between harmless bacteria and pathogenic species, allowing for precise detection and removal [5, 6].

Another emerging technology is the application of nanomaterials for capturing and isolating Mycoplasma and Acholeplasma from vegetable samples. Nanoparticles functionalized with specific ligands can selectively bind to the surface of these bacteria, facilitating their separation from the surrounding matrix. This approach offers high sensitivity and efficiency, enabling rapid and thorough removal of pathogens from vegetables [7, 8].

Furthermore, advancements in microfluidic devices and lab-on-a-chip systems have enabled the development of

portable and automated platforms for on-site detection and separation of bacterial contaminants. These miniaturized devices integrate sample preparation, analysis, and separation processes, offering a cost-effective and user-friendly solution for food safety monitoring. As research in the field of food microbiology continues to evolve, further innovations in separation techniques for Mycoplasma and Acholeplasma are expected. Collaborative efforts between scientists, food industry professionals, and regulatory agencies will be essential to implement these advancements into practical solutions for ensuring the safety of vegetable products [9, 10].

Conclusion

The separation of Mycoplasma and Acholeplasma from vegetables represents a critical aspect of food safety management. By leveraging the latest scientific and technological advancements, researchers are making significant strides in developing efficient and reliable methods for detecting and removing these pathogens from vegetable samples. These innovations not only protect consumer health but also contribute to the overall integrity and quality of the food supply chain.

References

1. Alam M, Hasan NA, Sadique A, et al. Seasonal cholera caused by *Vibrio cholerae* serogroups O1 and O139 in the coastal aquatic environment of Bangladesh. *Appl Environ Microbiol*. 2006;72(6):4096-104.
2. Alam M, Sultana M, Nair GB, et al. Toxigenic *Vibrio cholerae* in the aquatic environment of Mathbaria, Bangladesh. *Appl Environ Microbiol*. 2006;72(4):2849-55.
3. Gao MS, Azevedo NF, Wilks SA, et al. Persistence of *Helicobacter pylori* in heterotrophic drinking-water biofilms. *Appl. Environ. Microbiol*. 2008;74(19):5898-904.
4. Ramamurthy T, Yamasaki S, Takeda Y, et al. *Vibrio cholerae* O139 Bengal: Odyssey of a fortuitous variant. *Microbes and infection*. 2003;5(4):329-44.
5. Bhanumathi R, Sabeena F, Isac SR, et al. Molecular characterization of *Vibrio cholerae* O139 Bengal isolated from water and the aquatic plant *Eichhornia crassipes* in the River Ganga, Varanasi, India. *Appl Environ Microbiol*. 2003;69(4):2389-94.

*Correspondence to: Alexey Trushin, Department of Biophysics, Federal University, Russia, E-mail: trushin@alexey.ru

Received: 08-Mar-2024, Manuscript No. AAFMY-24-131247; Editor assigned: 09-Mar-2024, PreQC No. AAFMY-24-131247(PQ); Reviewed: 23-Mar-2024, QC No. AAFMY-24-131247; Revised: 28-Mar-2024, Manuscript No. AAFMY-24-131247(R); Published: 04-Apr-2024, DOI:10.35841/aaafmy-8.2.197

6. Mayer EA, Tillisch K, Bradesi S. Modulation of the brain–gut axis as a therapeutic approach in gastrointestinal disease. *Alime Pharmacol* 2006;24(6):919-33.
7. Hawkins KG, Casolaro C, Brown JA, et al. The microbiome and the gut–liver–brain axis for central nervous system clinical pharmacology: Challenges in specifying and integrating in vitro and in silico models. *Clinical Pharmacology & Therapeutics*. 2020;108(5):929-48.
8. Awouters F, Niemegeers CJ, Janssen PA. Pharmacology of antidiarrheal drugs. *Annu Rev Pharmacol Toxicol*. 1983;23(1):279-301.
9. Macke L, Schulz C, Koletzko L, et al. Systematic review: The effects of proton pump inhibitors on the microbiome of the digestive tract—evidence from next-generation sequencing studies. *Alimen Pharmacol Thera*. 2020;51(5):505-26.
10. Del Pozo-Acebo L, López de las Hazas MC, Margollés A, et al. Eating microRNAs: Pharmacological opportunities for cross-kingdom regulation and implications in host gene and gut microbiota modulation. *Briti J Pharmacol*. 2021;178(11):2218-45.