

# Advancing Genotyping Techniques: Tracking *Giardia lamblia* in Sohag Governorate, Egypt.

Vicky M Avery\*

Department of Morphology and Basic Pathology, Faculty of Medicine of Jundiaí, Jundiaí, Brazil

## Introduction

*Giardia lamblia*, a flagellated protozoan parasite, is a leading cause of gastrointestinal infections worldwide, particularly in regions with inadequate sanitation and hygiene practices. Sohag Governorate, Egypt, is no exception, facing its share of challenges in combating this microscopic foe. Understanding the genetic diversity of *G. lamblia* strains circulating in a specific region is crucial for effective disease management and control strategies. In this context, the application of advanced genotyping techniques like semi-nested PCR and restriction fragment length polymorphism (RFLP) has emerged as a valuable tool for molecular epidemiology studies.

Semi-nested PCR is a modification of conventional PCR that amplifies specific DNA fragments using two sets of primers. In the case of *G. lamblia* genotyping, this technique targets the small subunit ribosomal RNA (SSU rRNA) gene, a highly conserved region commonly used for species identification. The semi-nested approach enhances sensitivity and specificity by utilizing two rounds of PCR amplification, allowing for the detection of even low levels of parasite DNA.

Following PCR amplification, the next step involves RFLP analysis, which involves digesting the amplified DNA fragments with specific restriction enzymes. These enzymes cleave the DNA at specific recognition sites, resulting in fragments of varying lengths. By comparing the pattern of DNA fragments produced by different isolates, researchers can discern genetic variations among *G. lamblia* strains.

In Sohag Governorate, where *Giardia* infections pose a significant public health concern, the application of semi-nested PCR and RFLP techniques offers a promising avenue for understanding the genetic diversity of the parasite. By analyzing DNA samples obtained from infected individuals, researchers can identify distinct genotypes and assess their prevalence and distribution within the region.

Moreover, genotyping studies can provide insights into the transmission dynamics of *G. lamblia*, including the sources of infection and the potential for zoonotic transmission from animals to humans. This information is invaluable for designing targeted interventions aimed at interrupting the transmission cycle and reducing the burden of giardiasis in the community.

Despite its potential benefits, the application of genotyping techniques in resource-limited settings like Sohag Governorate may face challenges such as the availability of trained personnel, laboratory infrastructure, and financial resources. Addressing these challenges will require collaborative efforts involving local health authorities, academic institutions, and international partners to build capacity and support research initiatives in molecular epidemiology.

Furthermore, future studies should explore the utility of other molecular markers and advanced sequencing technologies to enhance the resolution and accuracy of genotyping data. Whole-genome sequencing, for example, can provide comprehensive insights into the genetic makeup of *G. lamblia* isolates, enabling more precise phylogenetic analysis and identification of genetic determinants associated with virulence and drug resistance.

## Conclusion

In conclusion, the genotyping of *G. lamblia* using semi-nested PCR and RFLP techniques represents a valuable approach for molecular epidemiology studies in Sohag Governorate, Egypt. By elucidating the genetic diversity and transmission dynamics of the parasite, these techniques contribute to the development of evidence-based strategies for the prevention and control of giardiasis. Moving forward, continued investment in research and capacity-building efforts is essential to harness the full potential of genotyping technologies in combating parasitic infections and safeguarding public health.

## References

1. Amoah AS, Boakye DA, Yazdanbakhsh M, et al. Influence of parasitic worm infections on allergy diagnosis in sub-Saharan Africa. *Curr Allergy Asthma Rep.* 2017;17:1-9.
2. Bethony J, Brooker S, Albonico M, et al. Soil-transmitted helminth infections: Ascariasis, trichuriasis, and hookworm. *Lancet.* 2006;367(9521):1521-32.
3. Crompton DW, Nesheim MC. Nutritional impact of intestinal helminthiasis during the human life cycle. *Annu Rev Nutr.* 2002;22(1):35-59.
4. Fletcher SM, Stark D, Harkness J, et al. Enteric protozoa in the developed world: a public health perspective. *Clin Microbiol Rev.* 2012;25(3):420-449.

\*Correspondence to: Vicky. M. Avery, Department of Morphology and Basic Pathology, Faculty of Medicine of Jundiaí, Jundiaí, Brazil, E-mail: V.Avery@griffith.edu.au

Received: 29-Dec-2023, Manuscript No. AAPDDT-24-135772; Editor assigned: 02-Jan-2024, PreQC No. AAPDDT-24-135772 (PQ); Reviewed: 17-Jan-2024, QC No. AAPDDT-24-135772; Revised: 22-Jan-2024, Manuscript No. AAPDDT-24-135772 (R); Published: 30-Jan-2024, DOI:10.35841/aapddt-9.2.182

5. Fotedar R, Stark D, Beebe N, et al. Laboratory diagnostic techniques for Entamoeba species. Clin Microbiol Rev. 2007;20(3):511-532.
6. Hotez PJ, Alvarado M, Basáñez MG, et al. The global burden of disease study 2010: interpretation and implications for the neglected tropical diseases. PLoS Negl Trop Dis. 2014;8(7):e2865.
7. Hotez PJ, Brindley PJ, Bethony JM, et al. Helminth infections: the great neglected tropical diseases. J Clin Invest. 2008;118(4):1311-21.
8. Petri Jr WA, Haque R. Entamoeba species, including amebic colitis and liver abscess. Mandell, Douglas, and Bennett's principles and practice of infectious diseases. 2015;3047-3058.
9. Pullan RL, Smith JL, Jasrasaria R, et al. Global numbers of infection and disease burden of soil transmitted helminth infections in 2010. Parasit Vectors. 2014;7:1-9.
10. Zumla A. Mandell, Douglas, and Bennett's principles and practice of infectious diseases. Lancet Infect Dis. 2010;10(5):303-4.