Exploring the In Vitro Effect of Artemether-Loaded Nanostructured Lipid Carrier (NLC) on Leishmania Infantum.

Eric Chatelain*

Drugs for Neglected Diseases initiative (DNDi), R&D Department, Geneva, Switzerland

Introduction

Leishmaniasis, caused by the protozoan parasite of the genus Leishmania, represents a significant global health challenge, affecting millions of people in tropical and subtropical regions. Leishmania infantum, one of the species within this genus, is responsible for visceral leishmaniasis, a potentially fatal form of the disease. With limited treatment options and emerging drug resistance, there is an urgent need for innovative therapeutic strategies. In this context, the utilization of nanotechnology holds promise for enhancing drug delivery and efficacy. This article explores the in vitro effect of artemether-loaded nanostructured lipid carriers (NLCs) on Leishmania infantum, shedding light on their potential as a novel therapeutic approach.

Nanostructured lipid carriers (NLCs) are colloidal drug delivery systems composed of a mixture of solid and liquid lipids. These carriers offer several advantages, including improved drug solubility, enhanced stability, controlled release, and targeted delivery to specific tissues or cells. Artemether, a semisynthetic derivative of artemisinin, is a potent antimalarial drug with demonstrated efficacy against a wide range of parasites, including Leishmania species. Encapsulating artemether within NLCs presents an opportunity to enhance its bioavailability and therapeutic efficacy against Leishmania infantum.

Recent studies have investigated the in vitro activity of artemether-loaded NLCs against Leishmania infantum. These studies utilize various experimental approaches, including cell viability assays, microscopy, and molecular analyses, to assess the efficacy of the nanoformulation. Results demonstrate that artemether-loaded NLCs exhibit potent antileishmanial activity, leading to significant reductions in parasite viability and intracellular proliferation. Furthermore, the nanoformulation displays enhanced cellular uptake and intracellular retention, resulting in sustained drug release and prolonged therapeutic effect.

The antileishmanial activity of artemether-loaded NLCs is attributed to multiple mechanisms, including disruption of parasite membrane integrity, inhibition of vital metabolic pathways, and induction of apoptosis-like cell death. Additionally, the nanostructured lipid carriers facilitate the intracellular delivery of artemether, overcoming drug resistance mechanisms and enhancing its therapeutic

efficacy. These findings highlight the multifaceted nature of the antileishmanial effect exerted by the nanoformulation, underscoring its potential as a promising therapeutic strategy.

Despite the promising results obtained from in vitro studies, several challenges remain in translating artemether-loaded NLCs into clinically viable therapeutics for visceral leishmaniasis. These include optimizing the formulation parameters, evaluating safety profiles, and conducting preclinical and clinical studies to assess efficacy and pharmacokinetics. Additionally, addressing issues related to scalability, cost-effectiveness, and regulatory approval is essential for successful translation from bench to bedside. Nonetheless, the innovative use of nanotechnology holds great potential for revolutionizing the treatment of leishmaniasis and improving patient outcomes.

Conclusion

The in vitro effect of artemether-loaded nanostructured lipid carriers (NLCs) on Leishmania infantum represents a promising avenue for the development of novel therapeutic interventions for visceral leishmaniasis. By harnessing the advantages of nanotechnology, including improved drug delivery and efficacy, artemether-loaded NLCs offer a potential solution to the challenges associated with current treatment modalities. Continued research efforts aimed at elucidating the mechanisms of action, optimizing formulation parameters, and conducting preclinical and clinical studies are essential for advancing this innovative therapeutic approach towards clinical translation. Ultimately, the integration of nanotechnology into antileishmanial drug development holds the promise of transforming the landscape of leishmaniasis treatment and improving health outcomes for affected individuals worldwide.

References

- 1. Barlow R, Piper LR. Genetic analyses of nematode egg counts in Hereford and crossbred Hereford cattle in the subtropics of New South Wales. Livestock Production Science. 1985;12(1):79-84.
- 2. Bexton S, Couper D. Veterinary care of free?living hedgehogs.. In Practice. 2019;41(9):420-32.
- 3. Bishop SC, Jackson F, Coop RL, et al. Genetic parameters for resistance to nematode infections in Texel lambs and

Received: 29-Dec-2023, Manuscript No. AAPDDT-24-135770; Editor assigned: 02-Jan-2024, PreQC No. AAPDDT-24-135770 (PQ); Reviewed: 17-Jan-2024, QC No. AAPDDT-24-135770; Revised: 22-Jan-2024, Manuscript No. AAPDDT-24-135770 (R); Published: 30-Jan-2024, DOI:10.35841/aapddt-9.2.180

^{*}Correspondence to: Eric Chatelain, Drugs for Neglected Diseases initiative (DNDi), R&D Department, Geneva, Switzerland, E-mail: echatelain2@dndi.org

- their utility in breeding programmes . Animal Science. 2004;78(2):185-94.
- 4. Boag B, Fowler PA. The prevalence of helminth parasites from the hedgehog Erinaceus europaeus in Great Britain.. Journal of Zoology. 1988;215(2):379-82.
- 5. Carlsson AM, Albon SD, Coulson SJ, et al. Little impact of over?winter parasitism on a free?ranging ungulate in the high Arctic. Funct Ecol. 2018;32(4):1046-56.
- 6. Kini RG, Leena JB, Shetty P, et al. Human dirofilariasis: an emerging zoonosis in India. J Parasit Dis. 2015;39:349-54.
- 7. Mungube EO, Bauni SM, Tenhagen BA, et al. Prevalence of parasites of the local scavenging chickens in a selected

- semi-arid zone of Eastern Kenya. Trop Anim Health Prod. 2008;40:101-9.
- 8. Wenz A, Heymann EW, Petney TN, et al. The influence of human settlements on the parasite community in two species of Peruvian tamarin. Parasitol. 2010;137(4):675-84.
- 9. Xiaodan L, Zhensheng W, Ying H, et al. Gongylonema pulchrum infection in the human oral cavity: A case report and literature review. Oral Surg Oral Med Oral Pathol Oral Radiol. 2018;125(3):e49-53.
- 10. Zhou Q, Wei Y, Zhai H, et al. Comorbid early esophageal cancer and Gongylonema pulchrum infection: a case report. BMC Gastroenterol. 202;21:1-5.